Nitric oxide (NO) is a redox-active molecule that plays a key role in a broad spectrum of physiological and developmental functions throughout the plant life cycle. Thus, NO is thought to contribute to the control of germination, leaf expansion, lateral root development, flowering, stomatal closure, and defenses against biotic and abiotic stresses (Neill et al., 2002; He et al., 2004; Hong et al., 2008; Wilson et al., 2008; Leitner et al., 2009). NO has also been implicated as a key regulator of senescence and leaf cell death in higher plants (Guo and Crawford, 2005; Zago et al., 2006). While descriptions of NO-mediated processes are accumulating, the details of how this small molecule regulates its target signaling networks remain largely unknown (Wang et al., 2010b). To date, numerous potential sources for NO production have been described in plants (Gupta et al., 2011), including a nitric oxide synthase (NOS)-like activity (Delledonne et al., 1998; Durner et al., 1998; Besson-Bard et al., 2008), nitrate reductase (NR; Stöhr and Stremlau, 2006; Stoimenova et al., 2007; Seligman et al., 2008), xanthine oxidoreductase (Corpas et al., 2008), and polyamine/hydroxylamine-mediated NO production (Rümer et al., 2009). Archetypal NOS enzymes found in animals (Palmer et al., 1988; Knowles and Moncada, 1994) and also the green alga Ostreococcus tauri (Foresi et al., 2010) catalyze the NADPH-dependent oxidation of l-Arg, producing L-citrulline and NO. A similar activity has been reported in higher plants, which is blunted by classical mammalian NOS inhibitors (Delledonne et al., 1998; Durner et al., 1998; Jasid et al., 2006; Vandelle and Delledonne, 2008; Corpas et al., 2009). However, no archetypal NOS-encoding genes have been identified so far in higher plants. Another potential source of NO production is NR, which catalyzes the production of this gas by the successive reduction of nitrite and nitrate. In Arabidopsis (Arabidopsis thaliana), NR is encoded by two genes, NIA1 and NIA2 (Yamasaki and Sakihama, 2000).
It has been shown that NO regulates a number of physiological processes directly by affecting gene transcription (Huang et al., 2002; Wang et al., 2002; Polverari et al., 2003; Parani et al., 2004; Shoulars et al., 2008). NO can also regulate specific physiological and developmental processes through its interplay with other small biomolecules. For example, NO can scavenge hydrogen peroxide (H$_2$O$_2$) and protects plant cells from damage (Beligni et al., 2002; Crawford and Guo, 2005). It was also reported that NO and H$_2$O$_2$ function in combination to elaborate cell death associated with the hypersensitive response (HR) following pathogen recognition (Delledonne et al., 2001). Furthermore, S-nitrosylation, the addition of an NO moiety to a Cys thiol to form an S-nitrosothiol (SNO), is emerging as a key redox-based posttranslational modification in plants and a pivotal mechanism to convey NO bioactivity. S-Nitrosylation may be integral to NO function during a variety of cellular processes. For example, S-nitrosylation of peroxiredoxin IIE during the defense response regulates the antioxidant function of this key enzyme and might contribute to the HR (Romero-Puertas et al., 2007). In animals, SNO formation has been reported to be involved in the regulation of apoptosis at multiple steps (Sawa et al., 1997; Hara et al., 2005; Mannick, 2007). In this context, the S-nitrosylation of GOSPEL (Sen et al., 2009), caspase 3 (Tsang et al., 2009), cyclooxygenase 2 (Carreras and Poderoso, 2007), FLICE (for FADD-like ICE) inhibitory protein (Chanvorachote et al., 2005), and apoptosis signal-regulating kinase1 (ASK1; Park et al., 2004) are important features of cell death in animals.

The global levels of S-nitrosylation are regulated by S-nitrosogluthathione reductase1 (GSNOR1) in Arabidopsis (Feechan et al., 2005). Loss-of-function mutations in GSNOR1 increased total cellular SNO content and compromised both nonhost and resistance (R) gene-mediated protection and also disabled basal defense responses (Feechan et al., 2005). Furthermore, this line was also perturbed in thermotolerance and responses to paraquat (Lee et al., 2008; Chen et al., 2009).

To generate insights into the potential role of S(NO) in the process of rice (Oryza sativa) leaf cell death and identify associated S-nitrosylated proteins, we conducted a forward genetic screen utilizing a large T-DNA-tagged population to identify mutants with perturbed (S)NO homeostasis. This approach uncovered the nitric oxide excess1 (noel) mutant, which accumulated more (S)NO in comparison with wild-type plants. Employing noel, we implicate NR-dependent NO production in rice leaf cell death in response to high light. Furthermore, our findings suggest a function for (S)NO and H$_2$O$_2$ in rice leaf cell death. Through the biotin-switch assay and nanoliquid chromatography-tandem mass spectrometry (LC-MS/MS), we identified S-nitrosylated proteins during cell death triggered by high light. These results provide initial insights into the mechanisms underpinning this process in rice leaves.

**RESULTS**

**Identification of noel**

In order to shed light on the role of (S)NO in leaf programmed cell death (PCD), we performed a large-scale screen to identify SNO content-altered mutants using the Saville-Griess assay from our T-DNA insertional mutant population (Ma et al., 2009). To simplify the assay with a large number of samples, we used 96-well plates and a Tecan Infinite M200 (Tecan Group) to undertake the photometrical measurement at 540 nm. Among the 380 screened mutants that show a PCD-like phenotype, 15 mutants with higher SNO content and four mutants with lower SNO content were selected for further analysis. One of them, designated noel, was identified because its SNO content was significantly higher than that of the wild type (Fig. 1A). No signs of damage were developed in the shaded parts of the leaves of noel grown in the field under natural growth conditions (high light; 1,600 pmol m$^{-2}$ s$^{-1}$) or under low light (400 pmol m$^{-2}$ s$^{-1}$; Supplemental Fig. S1, A and C). However, leaf damage, characterized by white variegated areas and cell death, developed when noel was cultivated under high light (Fig. 1, B and C; Supplemental Fig. S1B). When the 10-d-old low-light-grown noel plants were transferred to high light, the obvious leaf bleaching and cell death were developed in noel plants on the 2nd and 5th d, respectively (Supplemental Fig. S2A). Furthermore, no cell death was observed in noel plants cultivated under low light with different temperatures or photoperiods (Supplemental Fig. S2B). By contrast, exposure to high light together with different temperatures (20°C, 25°C, and 30°C) or photoperiods (light/dark: 6 h/18 h, 10 h/14 h, and 14 h/10 h) promoted cell death in all noel plants but not in wild-type controls (Supplemental Fig. S2C). These data implied that the observed cell death in noel plants is dependent on high light.

**Map-Based Cloning of NOE1/OsCATC**

Despite being identified from a T-DNA insertion population, the SNO overaccumulation and cell death phenotypes of noel did not cosegregate with a T-DNA insertion. Thus, the noel line without a T-DNA insertion was isolated by backcrossing and used for further analysis. An F2 mapping population was established using a cross between noel (Japanica) and Minghui 63 (indica). In the F2 population, we investigated 1,227 individual plants. Among them, 313 plants exhibited the noel mutant phenotype and 914 plants exhibited wild-type phenotypes, suggesting that the noel mutant was controlled by a single recessive gene. Rough mapping delimited the NOE1 locus at about 950 kb on chromosome 3 with genetic markers M2 and M8 (Fig. 1D). Further fine-mapping was performed using insertion-deletion molecular markers (Supplemental Table S1). The NOE1 locus
was narrowed down to a 61-kb region between MT19 and MT23 (Fig. 1D). After sequencing all 10 candidate genes within this region, a single-nucleotide G-to-T transition at the 168th position, which caused a Glu (GAG)-to-stop codon (TAG) change, was found in LOC_Os03g03910 of noe1. Subsequently, we identified another allelic mutant (confirmed by allelic test), noe1-2, with a similar aberrant phenotype in leaves and the same higher SNO content compared with the wild type. After sequencing the NOE1 (LOC_Os03g03910) locus, a 21-bp deletion was found, which led to a frame-shift mutation (Fig. 1D). NOE1 encodes the rice catalase domain-containing protein and has extremely high sequence similarity with catalase isozyme A (OsCATA) and catalase isozyme B (OsCATB) in rice and three catalase isozymes in Arabidopsis (Supplemental Fig. S3). As in the rice genome, there are only three catalase-encoding genes, so NOE1 should be OsCATC. Real-time quantitative PCR analysis revealed that the expression of OsCATA, OsCATB, and NOE1/OsCATC varied in different tissues (Fig. 1E). NOE1/OsCATC was mainly expressed in leaf blades, panicles, leaf sheaths, and culms, but expression was extremely low in roots. In contrast, OsCATA was expressed at a high level in leaf blades, panicles, and leaf sheaths but weakly expressed in roots and culms. As for OsCATB, high expression was observed in the panicles, leaf sheaths, and culms. In leaves, OsCATA and NOE1/OsCATC were highly expressed but OsCATB was much less abundant (Fig. 1E). In addition, no compensatory induction of OsCATA and OsCATB transcripts was observed in noe1 plants (Fig. 1F). These results implied that NOE1/OsCATC, but not OsCATA or OsCATB, was responsible for the redox homeostasis in leaves. To further validate NOE1/OsCATC, genetic complementation was carried out by introducing the entire cDNA of NOE1/OsCATC into noe1 plants. As expected, the leaf cell death phenotype was not observed in all noe1-C transgenic lines (Fig. 2A). Furthermore, enzyme activity assays showed that the catalase activities of complemented plants were also restored to wild-type levels (Fig. 2B). Consequently, H₂O₂ contents in noe1-C plants were also decreased to that of the wild type (Fig. 2C) as well as the content of SNO (Fig. 2D). Consistent with the impairment of NOE1/OsCATC, total catalase activity in noe1 plants was reduced dramatically to about 30% of that of the wild type (Fig. 2B), and more H₂O₂ and cell death accumulated (Fig. 2C; Supplemental Fig. S4). After the addition of 200 μM CAT (an H₂O₂ scavenger), the leaf cell death in noe1 plants did not develop at the 2nd or 5th d under high light (Supplemental Fig. S5, A and C). Furthermore, cell death in noe1 suspension cells was reduced from 53.9% to 16.6% after treatment with 200 μM CAT (Supplemental Fig. S5E). These data suggest that leaf cell death in noe1 may be caused by H₂O₂ overaccumulation. Taken together, loss-of-function mutations in NOE1/OsCATC are responsible for the phenotypes displayed in the noe1 mutants.
H2O2 Stimulates NO Production

Labeling NO with the cell-permeable, NO-specific fluorescent probe 4-amino-5-methylamino-2',7'-difuorofluorescin diacetate (DAF-FM DA) and imaging by confocal microscopy revealed that the NO content in noe1 suspension cells was about 2.3-fold greater than that of the wild type (Fig. 3, A and B). To elucidate the relationship between H2O2 and NO, treatment of wild-type and noe1 suspension cells with 1 mM H2O2 was performed, and DAF-FM DA analysis showed that H2O2 induced NO production significantly (Fig. 3, A and B). The seedlings of wild-type and noe1, and noe1-C plants grown under high light (1,600 μmol m⁻² s⁻¹). These data were obtained from three independent replicates. FW, Fresh weight.

H2O2-Induced NR-Dependent Generation of NO

To elucidate the main source responsible for NO production in noe1 plants under high light, the activities of NOS and NR were measured. No difference in NOS activity was observed between wild-type and noe1 plants (Fig. 4A). NR activity in noe1 plants, however, was approximately twice that of the wild type (Fig. 4B). Quantitative real-time PCR analysis revealed that the expression of OsNIA1 and OsNIA2 increased 3- and 9-fold, respectively, in noe1 plants (Fig. 4, C and D), in line with the increase of NO activity. As a result, the content of the NR substrate, nitrate, was decreased in noe1 plants (Fig. 4E). To further confirm that NO was mainly produced from NR activity, tungstate (a NR inhibitor) and N⁶-monomethyl-L-arginine, monoa Ace tate salt (L-NMMA; an NOS inhibitor) were used, and SNO contents were monitored. The content of SNOs was inhibited in a dose-dependent manner when noe1 seedlings were treated with tungstate, while no change in SNO content was observed in noe1 seedlings treated with L-NMMA (Fig. 4F). These results implied that the excess NO in noe1 was mainly generated from NR. Presumably, this increased NO concentration subsequently resulted in greater SNO levels.

NO Is Required for H2O2-Induced Leaf Cell Death

To identify the role of NO in H2O2-induced leaf cell death modulated by light (Supplemental Figs. S1 and S2), treatments of wild-type and noe1 seedlings with PTIO and sodium nitroprusside (SNP; an NO donor), both alone and in combination, were carried out. After transferring the 10-d-old low-light-grown noe1 plants to high light, obvious leaf bleaching and cell death formation occurred on the 2nd and 5th d, respectively (Fig. 5, A and C). The addition of 200 μM PTIO, which depletes NO, resulted in significantly reduced leaf cell death (Fig. 5). By contrast, leaf cell death in noe1 plants was promoted following treatment with 2.5 mM SNP (Fig. 5). Cell death could also be triggered in wild-type plants following the addition of 2.5 mM SNP for 5 d (Fig. 5, C and D), demonstrating a requirement for NO in the process of cell death. Similar treatments were performed with dark-grown rice suspension cells, in which cell death can be accurately measured. The results showed that the relative cell death ratio in noe1 suspension cells increased from 14.2% to 54.5% when exposed to low light (400 μmol m⁻² s⁻¹) for 1 d (Fig. 5E). After the addition of 200 μM PTIO, the ratio of cell death was reduced to 19.7% when exposed to low light for 1 d, and this level was similar to that found in wild-type cells (15.4%). When cultured with 2.5 mM SNP.

Figure 2. Phenotypes of noe1 were complemented by NOE1/OsCATC. A, Phenotypic comparison among 3-month-old wild-type (WT), noe1, and complemented transgenic noe1-C plants. B to D, Total extractable leaf catalase activities and H2O2 and SNO contents in 3-month-old wild type, noe1, and noe1-C plants grown under high light (1,600 μmol m⁻² s⁻¹). These data were obtained from three independent replicates. FW, Fresh weight.

Contrast, H2O2 content was not altered after treatment of high-light-grown noe1 plants with 2-phenyl-4,4,5,5-tetramethylimidazoline-1-oxyl 3-oxide (PTIO; an NO scavenger; Fig. 3F). These results suggested that H2O2-stimulated NO production may cause the increased SNO levels in leaves of noe1 plants under high light.
under low light for 1 d, the relative cell death ratio in wild-type and noe1 suspension cells increased to 42.4% and 73.1%, respectively, significantly greater than control values (wild type and noe1: 15.0% and 54.5%, respectively; Fig. 5E). These results suggest that NO is a pivotal mediator for H2O2-induced leaf cell death in rice.

Protein S-Nitrosylation Is Involved in H2O2-Induced Leaf Cell Death

It is known that protein S-nitrosylation is a fundamental mechanism to transduce NO bioactivity (Bessonnard et al., 2008) and that GSNOR regulates global levels of S-nitrosylation (Feechan et al., 2005). OsGSNOR is encoded by a single gene in rice that has very high sequence similarity with AtGSNOR (Supplemental Fig. S7). To investigate the potential role of OsGSNOR in leaf cell death, OsGSNOR overexpression (GSNOR-O noe1) or knockdown (GSNOR-R noe1) transgenic plants in the noe1 background were generated (Fig. 6A). GSNOR-O noe1 plants with increased GSNOR activity and GSNOR-R noe1 plants with reduced GSNOR activity were selected for further analysis (Fig. 6B). Employing the Saville-Griess assay, lower SNO content was found in GSNOR-O noe1 plants relative to noe1 lines. In contrast, GSNOR-R noe1 plants exhibited an increased level of these redox molecules (Fig. 6C). These data are consistent with those from Arabidopsis, which suggested that GSNOR controls the total level of SNOs in cells (Feechan et al., 2005). The biotin-switch assay, labeling of S-nitrosylated proteins with a biotin moiety specifically on S-nitrosylated Cys residues (Jaffrey and Snyder, 2001), further confirmed these results (Fig. 6D). When cultivated under high light, there was no difference in phenotype between either the GSNOR-O noe1 or the GSNOR-R noe1 transgenic line relative to that of noe1 plants (data not shown). However, when these lines were grown under low light for 10 d and then subsequently exposed to sustained H2O2 synthesis (generated by the Glc/Glc oxidase system, which could moderately release H2O2) for 3 d, leaf cell death in GSNOR-O noe1 plants was less than that observed in noe1 plants (Fig. 6E). Collectively, these findings imply that protein S-nitrosylation is involved in H2O2-induced leaf cell death.

Proteomic Identification of S-Nitrosylated Proteins

To identify possible candidates for S-nitrosylation integral to H2O2-induced leaf cell death, the biotin-switch assay and LC-MS/MS were performed. Total protein, extracted from wild type and noe1 leaves grown under low light for 10 d and subsequently transferred to high light for 2 d, were blocked with methyl methanethiosulfonate and labeled with N-[6-(biontaminadobexyl)-3‘-(2’-pyridyldithio)propionamide (biotin-HPDP) (Wang et al., 2009); after precipitation with acetone, the resuspended mixture was first digested with trypsin before purification with affinity chromatography to harvest the biotin-labeled peptides (biotin-HPDP) (Wang et al., 2009); after precipitation with acetone, the resuspended mixture was first digested with trypsin before purification with affinity chromatography to harvest the biotin-labeled peptides (biotin-HPDP) (Wang et al., 2009).
tion processing, and redox regulation (Fig. 7B; Supplemental Fig. S9). Almost half the proteins identified were reported previously as being subject to S-nitrosylation (Supplemental Tables S7–S9), indicating the reliability of these results. Among the proteins identified in noe1 plants, approximately 37.5% functioned in general metabolism and 21% in genetic information processing (Fig. 7B). Proteins related to environmental adaptation and redox homeostasis accounted for 10% and 14%, respectively (Fig. 7B). Among the seven redox-related S-nitrosylated proteins identified in noe1, glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and thioredoxin (TRX) were previously reported to be involved in cell death in animals, and their functions were linked with their S-nitrosylation (Sumbayev, 2003; Hara et al., 2005; Table I). These results further implied that protein S-nitrosylation is involved in H2O2-induced leaf cell death.

Figure 4. NR was mainly responsible for the S (NO) accumulation in noe1. A to E, NOS (A) and NR (B) activities, relative expression levels of OsNIA1 (C) and OsNIA2 (D) by quantitative RT-PCR (with the wild-type value [WT] set as 1.0), and nitrate content (E) in wild-type and noe1 plants. FW, Fresh weight. F, Effects of tungstate (an NR inhibitor) and l-NMMA (an NOS inhibitor) on endogenous NO production in wild-type and noe1 plants. Both 10-d-old wild-type and noe1 seedlings cultivated under low light (400 μmol m⁻² s⁻¹) were treated with tungstate or l-NMMA under high light (1,600 μmol m⁻² s⁻¹) for 2 d. These data were obtained from three independent replicates. Tungstate100 and Tungstate200, 100 and 200 μM tungstate; l-NMMA200 and l-NMMA300, 200 and 300 μM l-NMMA; PTIO200, 200 μM PTIO. * P < 0.05, ** P < 0.01 (t test).

Figure 5. NO was required for leaf cell death. A to D, Both 10-d-old wild type (WT) and noe1 seedlings cultivated under low light (400 μmol m⁻² s⁻¹) were transferred to high light (1,600 μmol m⁻² s⁻¹). Leaf symptoms and relative cell death (percentage of total leaf area) are given for wild-type and noe1 seedlings induced by the addition of 200 μM PTIO (an NO scavenger) and 2.5 mM SNP (an NO donor) for 2 d after treatment (DAT; A and B) and 5 DAT (C and D) under high light. E, Effects of 200 μM PTIO and 2.5 mM SNP on relative cell death of wild-type and noe1 suspension cell cultures. These data were obtained from three independent replicates.
DISCUSSION

NO and reactive oxygen species (ROS) synthesis are routine requirements for plant cells to undergo PCD; however, in some contexts, evidence has been presented that either can induce HR independently of the other (Clarke et al., 2000). The emerging data also suggest that significant cross talk occurs between NO and ROS (Delledonne et al., 2001), although the molecular details of this interchange remain undefined. Here, we identified a rice NO accumulation mutant, noe1, following a map-based cloning approach. NOE1 was found to encode the rice catalase OsCATC. Further analysis revealed that NOE1/OsCATC is the ortholog of AtCAT2 (Iwamoto et al., 2000; Zimmermann et al., 2006). A previous report showed that cell death in cat2 plants grown in long days was linked to photoperiod, not light intensity (Queval et al., 2007). However, these data contrasted with our findings with noe1 plants. When exposed to high light, the lack of leaf catalase activity in noe1 plants resulted in an accumulation of H2O2, which further induced NO production. Conversely, the reduction of intracellular NO levels using the scavenger PTIO, or the levels of SNO by overexpression of OsGSNOR, alleviated leaf cell death triggered by H2O2 in noe1 plants. In addition, treatment of noe1 plants and OsGSNOR RNA interference (RNAi) transgenic lines with the NO donor SNP promoted cell death. Taken together, these results imply that NO and SNO are important mediators of light-dependent leaf cell death in rice. Previous work has also shown that NO acts as...
a pivotal positive mediator in Cd²⁺-induced PCD in suspension cell cultures (De Michele et al., 2009), leaf senescence (Carimi et al., 2005), and the HR (Zaninotto et al., 2006). Additionally, cell death in dicotyledonous soybean (Glycine max) suspension cultures has been reported to be regulated by the intracellular NO-ROS ratio (Delledonne et al., 2001). In contrast, it was suggested that NO acts as an antagonist to delay PCD in barley (Hordeum vulgare) aleurone layers and leaf senescence in Arabidopsis (Beligni et al., 2002; Guo and Crawford, 2005; Mishina et al., 2007). These discrepancies in the role of NO in cell death might be due to the differences in plant species, redox state, and growth conditions.

The molecular mechanism(s) of NO generation in response to various stimuli still remain controversial. However, the addition of tungstate (an NR inhibitor) dramatically blunted NO accumulation in rice following exposure to high light, suggesting that increased NO accumulation in noe1 plants in response to this stimulus was predominantly generated from NR. Moreover, the increase of NR activity in noe1 plants under light high correlated with the up-regulation of two NR-encoding genes, OsNIA1 and OsNIA2. In Arabidopsis, increased NR activity in response to light, nitrate, and Suc was also congruent with increased expression of NIA1 and NIA2 (Cheng et al., 1992; Campbell, 1999). Thus, these observations provide further circumstantial evidence of a role for NR in NO synthesis in response to high light. Interestingly, promoter analysis also revealed a 28-bp motif in the promoter region of OsNIA1 and OsNIA2, which is similar to the redox-responsive motif identified in OsSODCC1, OsTRXH, and OsGRX (Tsukamoto et al., 2005). Thus, increasing H₂O₂ levels following exposure to high light might increase NR expression through this redox-responsive element. Furthermore, several reports have suggested that NR activity can be modulated at the posttranscriptional level. For example, H₂O₂ is required for the activation of mitogen-activated protein kinase (MAPK) 6, which can subsequently increase NR activity by phosphorylation of NIA2 at Ser-627 (Wang et al., 2010a). Also, NIA2 can be phosphorylated by a calcium-dependent protein kinase at Ser-534 in the hinge 1 region in response to light and other environmental signals (Su et al., 1996). Therefore, increased cellular H₂O₂ concentrations following exposure to high light might also increase NR activity through the phosphorylation of either Ser-627 or Ser-534.

As S-nitrosylation is a major route for the transduction of NO bioactivity, we identified 73 and 100 S-nitrosylated proteins from wild type and noe1 plants, respectively, by employing the biotin-switch assay and LC-MS/MS. Among these proteins, there were 52 S-nitrosylated proteins from both wild-type and noe1 plants. These proteins are involved in a wide array of cellular activities, such as genetic information processing, general metabolism, environmental adaptation, and redox homeostasis. Both GAPDH and TRX, which were reported to be involved in animal cell death (Sumbayev, 2003; Hara et al., 2005), were only S-nitrosylated in noe1 plants (Table I). In mammals, GADPH is S-nitrosylated at its active site, Cys-150, which promotes its association with the E3 ligase, Siah1, resulting in nuclear translocation of the SNO-GADPH-Siah1 complex (Mannick,

![Figure 7.](Image)

**Figure 7.** Identification of S-nitrosylated proteins from wild-type (WT) plants and the noe1 mutant using LC-MS/MS. A, Number of S-nitrosylated proteins in wild-type and noe1 plants. B, Functional categorization of S-nitrosylated proteins identified only from noe1 plants. Both 10-d-old wild-type and noe1 seedlings cultivated under low light (400 μmol m⁻² s⁻¹) were transferred to high light (1,600 μmol m⁻² s⁻¹) for 2 d before sample collection.
Within the nucleus, SNO-GAPDH stabilizes Siah1 and facilitates the ubiquitination and degradation of nuclear proteins (Hara et al., 2005), which promotes cell death (Foster et al., 2009; Tristan et al., 2011). GAPDHs also play a central role in the carbon economy of plant cells, and higher plants possess four distinct isoforms of this protein (Hajirezaei et al., 2006; Muñoz-Bertomeu et al., 2009, 2010). A knockout line of cytosolic GAPDH shows reduced levels of oxygen uptake and ATP but increased ROS accumulation and also a higher density of trichomes (Rius et al., 2006). TRX is a key modulator of redox status (Vieira Dos Santos and Rey, 2006). During nitrosative stress, S-nitrosylation of TRX, presumed to be S-nitrosylated at Cys-32 or Cys-35 within the active site, promotes apoptosis, presumably by inhibiting the oxidoreductase function of this enzyme and by facilitating the release of sequestered ASK1 (Sumbayev, 2003; Hess et al., 2005). ASK1 is a MAPK that specifically activates the Jun N-terminal kinase and p38 MAPK signaling networks and is integral to tissue necrosis factor-α-induced apoptosis (Hess et al., 2005). Thus, S-nitrosylation of both GAPDH and TRX in noe1 but not in wild-type plants parallels the development of cell death in animal systems.

Ascorbate peroxidase (Vacca et al., 2004), glutaredoxin (Saeed et al., 2010), acyl carrier protein (Feng et al., 2009), phosphomannomutase (Hoeberichts et al., 2004), Fru-bisP aldolase isozyme (Yao et al., 2004), triose phosphate isomerase (Gnerer et al., 2006), pyruvate kinase (Seták et al., 2007), acidic 27-kD endochitinase (Shin et al., 2009), and lactate/malate dehydrogenase (Matthews et al., 2004) have all previously been reported to be involved in programmed cellular execution (Table I; Supplemental Table S4). However, the molecular mechanisms associated with the functions of these proteins in cell death remain to be established. Our findings imply that all of these proteins may also be S-nitrosylated in the rice noe1 mutant. Interestingly, although noe1 plants were not subjected to any pathogen challenge, a pathogenesis-related Bet v I family protein and PR1b were both S-nitrosylated (Supplemental Table S5), suggesting that this modification might regulate the activity of these gene products. Ribosome-related proteins (Supplemental Table S6) were also S-nitrosylated, indicating that this modification might also be involved in the regulation of protein translation. Furthermore, an elongation factor was also found to be a target for S-nitrosylation, suggesting that NO may control protein synthesis in response to H$_2$O$_2$ (Shenton and Grant, 2003; Lindermayr et al., 2005). Despite a central role for GSNOR in SNO homeostasis, information on the regulation of this enzyme is limited (Besson-Bard et al., 2008). Constitutively increased GSNOR activity in Arabidopsis atgsnor1-1 and atgsnor1-2 mutants correlated with elevated AtGSNOR1 mRNA levels (Feechan et al., 2005). However, GSNOR protein abundance, rather than the magnitude of transcripts, was promoted by paraquat (Chen et al., 2009). In contrast, neither atgsnor1-1 nor transgenic lines (Supplemental Fig. S8), suggesting that NO might function downstream of H$_2$O$_2$ in light-driven leaf cell death in rice.

Collectively, we have identified an NO accumulation mutant, noe1, in rice. Map-based cloning revealed that NOE1 encoded OsCATC. noe1 resulted in an increase of H$_2$O$_2$ in the leaves, which consequently promoted NO production via the activation of NR.

Figure 8. A hypothetical model for the role of NO and SNOs in H$_2$O$_2$-induced leaf cell death in rice. RNS, Reactive nitrogen species; -SH, free thiol.
Our data imply that NO and SNO are important mediators of light-dependent leaf cell death in rice. Furthermore, we have uncovered a series of S-nitrosylated proteins in both noe1 and wild-type plants, which may help to reveal the role of this posttranslational modification in the control of light-mediated leaf cell death in rice (Fig. 8).

MATERIALS AND METHODS

Plant Growth and Sampling

Rice (Oryza sativa) plants were cultivated in the experimental field of the Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, in Beijing. Rice seeds were performed using 10-d-old rice seedlings. Seeds of the wild type and noe1 were submerged in water for 2 d at 37°C, then the uniformly germinated seeds were cultured on 96-well plates supplied with half-strength Murashige and Skoog basal medium (pH 5.8) for 10 d in the growth chamber at 30°C for 14 h (low light, 400 μmol m−2 s−1) and 28°C for 10 h (dark). After the seedlings were transferred to high light (1,600 μmol m−2 s−1), leaf samples were taken on the 2nd and 5th d to measure the NO and H2O2 contents. The chemicals used to treat the 10-d-old seedlings, SNP (an NO donor), PTIO (an NO scavenger), CAT (an H2O2 scavenger), tungstate (an NR inhibitor), and -NMA (an NOS inhibitor) were added with 200 mL of half-strength Murashige and Skoog medium. All chemicals were purchased from Sigma-Aldrich. GSNOR-O noe1 and GSNOR-R noe1 transgenic lines and controls were cultivated under low light for 10 d and treated with 200 μm SNP, and H2O2 (about 400 μm) and L-NMMA (an NOS inhibitor) and PTIO, 2.5 mM SNP, and H2O2 (about 400 μm) with 1% polyvinylpyrrolidone and centrifuged at 2,500 g fresh weight) were homogenized in 2 mL of phosphate buffer (pH 7.8) with some modifications (Qian et al., 2010). Briefly, leaves were submerged in water for 2 d at 37°C, then the NOE1 callus was transformed by an Agrobacterium-mediated method as described previously (Liu et al., 2007). The primer sequences used for vector construction are listed in Supplemental Table S3.

Cell Death Analysis

Relative cell death of cell suspension was assayed as described previously with some modifications (Delledonne et al., 2001). Briefly, after 24-h treatments with CAT, PTIO, SNP, and SNP+PTIO under low light (400 μmol m−2 s−1), the rice suspension cells were stained with 0.05% Evans blue (Sigma-Aldrich) for 15 min. The excess dye was removed by extensive washing, and dye bound to dead cells was solubilized in 50% methanol/1% SDS in a volume of 20 mL for 30 min at 50°C and quantified by optical density at 595 nm (OD595). The OD595 values of rice cell suspensions saturated with methanol for 15 min was taken as the total killed cell death. The relative cell death is expressed as an OD595 ratio of different treatments to total killed cell death. The relative cell death of wild-type and noe1 seedlings leaves was analyzed using Adobe Photoshop CS and expressed as a percentage of cell death region to total leaf region.

Histochemical Detection

H2O2 was detected by 3,3′-diaminobenzidine staining as described previously with some modifications (Thordal Christensen et al., 1997). The fully expanded leaves of 2-month-old wild-type and noe1 plants were detached and infiltrated with the 0.1% 3,3′-diaminobenzidine solution. The sampled leaves were placed in a growth chamber for 5 h at 28°C and cleared in boiling ethanol (95%) for 10 min. The chlorophyll was removed by incubating in 95% ethanol overnight before photographing.

Malondialdehyde Content Assay

Malondialdehyde (MDA) was regarded as an indicator of lipid peroxidation and cell death. The content of MDA was determined with a MDA assay kit (Beyotime) with some modifications (Qian et al., 2010). Briefly, leaves (0.02 g fresh weight) were homogenized in 2 mL of phosphate buffer (pH 7.8) containing 1% polyvinylpyrrolidone and centrifuged at 2,500g for 10 min. MDA in the supernatant was determined according to the reaction with thiobarbituric acid and measured the absorbance at 450, 532, and 600 nm. According to the equation 6.45 × (OD532 − OD600) − 0.56 × OD450 the MDA content was expressed as μM MDA g−1 fresh weight.

Quantitative Measurement of H2O2 Content and Catalase Activity

The measurement of H2O2 production was performed by extracting H2O2 from leaves according to a previously described method (Huang et al., 2009) and quantifying the molecule with the Amplex Red Hydrogen Peroxidase Assay Kit (Molecular Probes, Invitrogen) according to the manufacturer’s instructions. Catalase activity was carried out with a Catalase Assay Kit (Beyotime) according to the manufacturer’s instructions.
Neutralization of Ion Leakage

The measurement of ion leakage was performed according to a previously described method (Woo et al., 2001). Five different leaf discs from wild-type and noe1 plants were placed into a 100-mL beaker containing 40 mL of distilled deionized water. After shaking at 120 rpm for 3 h, the conductivity (C1) was measured. Then, the discs were boiled for 10 min and shaken for 1 h, and the conductivity (C2) was measured again. The ion leakage was calculated according to the equation (C1/C2) × 100%.

Saville-Griess Assay

SNO content was measured using the Saville-Griess assay as described with minor modifications (Feechan et al., 2005). In the Saville-Griess assay, S-NO bonds were broken by mercurochloride, and the released NO reacted with sulfanilamide. The resulting diazonium salt is coupled with the aromatic amine N-(1-naphthyl)-ethylenediamine to form an intensely colored azo dye that can be measured at 540 nm (Feechan et al., 2005; King et al., 2005). Briefly, fine powder of plant tissues from liquid nitrogen was lysed in 600 μL of extraction buffer (50 mM Tris-HCl, pH 8.0, and 130 mM NaCl) containing 1 mM protease inhibitor phenylmethylsulfonyl fluoride (PMSF) and incubated on ice for 20 min. After centrifugation at 10,000 rpm for 15 min at 4°C, 160 μL of supernatant clear cell lysate was incubated with the same volume of 1% sulfanilamide and 0.1% N-(1-naphthyl)-ethylenediamine with or without the addition of 3.75 mM HgCl2 for 20 min in the dark. Then, SNO content was measured photometrically at 540 nm using 96-well plates with Tecan Infinite M200 (Tecan Group). The SNO content was calculated according to the absorption at 540 nm and a GSNO concentration standard curve (Feechan et al., 2005).

Determination of Endogenous NO Content of Suspension Cells with DAF-FM DA

The endogenous NO level of suspension cells was detected by imaging the cell-permeable, NO-specific fluorescent probe DAF-FM DA (Invitrogen) by confocal microscopy with some modifications (Zha et al., 2007). Wild-type and noe1 suspension cells (untreated or treated with 1 mM H2O2 for 30 min at 28°C) were incubated with 100 μM Tris-HCl (pH 7.6) containing 5 μM DAF-FM DA at 28°C for 40 min. The cells were then washed three times with fresh Tris-HCl (100 μM, pH 7.6) to remove excess probe and incubated for an additional 20 min to allow complete destetization of the intracellular dicarboxates. The incubated cells were visualized using a laser confocal scanning microscope (Leica TCS SPS). Excitation/emission wavelengths were 488/515 nm. Data are presented as mean pixel intensities. Fifty to 100 suspension cells were observed per treatment for three independent replicates.

Determination of NOS and NR Activities and Nitrate Content

The activities of NOS and NR were determined with the NOS Assay Kit and the Nitrate/Nitrite Assay Kit (Beyotime) with some modifications (Xiong et al., 2009). Total protein was extracted using buffer containing 100 mM HEPES-KOH (pH 7.5), 1 mM EDTA, 10% glycerol, 5 mM dithiothreitol, 0.1% Triton X-100, 0.5 mM PMSF, 25 μM leupeptin, 5 μM Na2MoO4, and 1% p-acyloniaprymolidine. The reaction was started by adding GSNO (Alexis Biochemicals) at a final concentration of 400 μM.

Protein Extractions

Protein extractions were performed according to a previously described method (Jaffrey and Snyder, 2001). Briefly, leaf tissues (200 mg) were ground in liquid nitrogen using a mortar and pestle. Soluble proteins were extracted from the resulting powder at 4°C in 1 mL of HEN buffer (250 mM HEPES-NaOH, pH 7.7, 1 mM EDTA, 0.1 mM neocuproine, and 10 mM PMSF) in the dark. The total extracts were incubated on ice for 20 min and centrifuged at 10,000 rpm for 15 min at 4°C.

Biotin-Switch Assay and Isolation of Biotinylated Proteins

The biotin-switch assay was performed as described previously (Wang et al., 2009). Three milligrams of soluble total protein was used for the assay. After the biotin-HDPP labeling step, around 30 μg of labeled soluble protein was used for western blot according to the standard protocol. Two volumes of −20°C acetone were added to the rest of the samples, incubated for 20 min at −20°C, and then centrifuged at a minimum of 2,000g for 10 min at 4°C. The supernatant was discarded to remove biotin-HDPP, then the wash of the tube and the surface of the pellets were gently rinsed with −20°C acetone to remove traces of biotin-HDPP. The pellets were resuspended in 0.1 mL of 25 mM NH4HCO3 buffer per mg of protein in the initial protein sample and digested with trypsin (sequencing-grade modified trypsin; Promega) with a protease:protein ratio of 1:20 (w/v) at 4°C overnight. Two volumes of neutralization buffer (200 mM HEPES-NaOH, pH 7.7, 100 mM NaCl, 1 mM EDTA, and 0.5% Triton X-100) was added, then 15 μL of packed streptavidin-agarse resin per mg of protein was used in the initial protein sample to purify biotinylated proteins. The proteins were incubated with resin for 1 h at room temperature with continuous shaking, and the resin was washed five times with 10 volumes of neutralization buffer plus 600 mM NaCl and centrifuged at 200g for 5 s at room temperature between each wash. The resin was incubated with elution buffer (20 mM HEPES-NaOH, pH 7.7, 100 mM NaCl, 1 mM EDTA, and 100 mM mercaptoethanol) to recover the bound proteins. Purified protein extracts were identified by LC-MS/MS analysis to find S-nitrosylated proteins.

According to a previous report (Wang et al., 2009), LC-MS/MS experiments were carried out on a Thermo LTQ Orbitrap XL instrument (Thermo Electron) equipped with a house-made source and an Eksigent two-dimensional nanolC system (Eksigent Technologies). Sample analyses were performed using an in-house-made reverse-phase C18 capillary column (15 cm × 100 μm i.d.). The peptides were sequentially eluted with a gradient of 0% to 80% buffer B (acetonitrile with 0.5% formic acid) in buffer A (water with 0.5% formic acid) at a flow rate of 400 nL min−1. The electrospray source parameters were 3.1 kV of electrospray voltage, the range of mass-to-charge ratio was from 300 to 1,800, and the normalized collision energy of MS/MS was 35%. The data were analyzed with Bioworks 3.31 and Sequest 2.8, and the results were filtered by Xcor (the cross-correlation value from the search) +1 > 1.9, +2 > 2.5, +3 > 3.75, sp > 500, Δm (the Δm correlation value) > 0.1, Rsp ≤ 5. The pathways of proteins that exert their roles were performed by searching in Kyoto Encyclopedia of Genes and Genomes (http://www.genome.jp/dbget-bin/www_bfind?O.sativa).

Supplemental Data

The following materials are available in the online version of this article.

Supplemental Figure S1. Leaf cell death in noe1 is dependent on high light.

Supplemental Figure S2. Leaf cell death in noe1 is dependent on light intensity.

Supplemental Figure S3. NOE1/OsCATC sequence alignment with other catalase proteins from rice and Arabidopsis.

Supplemental Figure S4. More H2O2 accumulated and cell death occurred in noe1.

Supplemental Figure S5. H2O2-induced leaf cell death in noe1.

Supplemental Figure S6. H2O2 and SNO levels in wild-type and noe1 seedlings at different time points.

Supplemental Figure S7. Sequence alignment of GSNOR from rice and Arabidopsis.
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LITERATURE CITED


Guo FQ, Crawford NM (2005) Arabidopsis nitric oxide synthase1 is targeted to mitochondria and protects against oxidative damage and dark-induced senescence. Plant Cell 17: 3436–3450


Primers used for vector construction.

Supplemental Table S3.

Supplemental Table S4.

Supplemental Table S5.

Supplemental Table S6.

Supplemental Table S7.

Supplemental Table S8.

Supplemental Table S9.

Supplemental Figure S8.

Supplemental Figure S9.

Supplemental Table S1.

Primers used for cloning.

Supplemental Table S2.

Primers used for quantitative RT-PCR.

Supplemental Figure S10.


